

Synthesis and Application of Fe₃O₄@Au Composite Nanoparticles as Magnetic Resonance/Computed Tomography Dual-Modality Contrast Agent

Abstract

Background: None of the molecular imaging modalities can produce imaging with both anatomical and functional information. In recent years, to overcome these limitations multimodality molecular imaging or combination of two imaging modalities can provide anatomical and pathological information. **Methods:** Magnetic iron oxide nanoparticles were prepared by co-precipitation method and then were coated with silica according to Stober method. Consequently, silica-coated nanoparticles were amino-functionalized. Finally, gold nanoparticles assembled onto the surfaces of the previous product. Cytotoxicity effects of prepared Fe₃O₄@Au nanoparticles were evaluated by 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide assay on human hepatocellular carcinoma cells. Their ability as a dual-mode contrast agent was investigated by magnetic resonance (MR) and computed tomography (CT) imaging. **Results:** Fe₃O₄@Au nanoparticles were spherical undersize of 75 nm. X-ray diffraction analysis confirmed the formation of Fe₃O₄@Au nanoparticles. The magnetometry result confirmed the superparamagnetism property of prepared nanoparticles, and the saturation magnetization (M_s) was found to be 33 emu/g. Fe₃O₄@Au nanoparticles showed good cytocompatibility up to 60 µg/mL. The results showed that the Fe₃O₄@Au nanoparticles have good r_2 relaxivity (135.26 mM⁻¹s⁻¹) and good X-ray attenuation property. **Conclusion:** These findings represent that prepared Fe₃O₄@Au nanoparticles in an easy and relatively low-cost manner have promising potential as a novel contrast agent for dual-modality of MR/CT imaging.

Keywords: *Computed tomography, gold nanoparticles, iron oxide nanoparticles, magnetic resonance imaging*

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Introduction

Molecular imaging can be defined as the imaging of targeted macromolecules and biological processes in living organisms.^[1,2] Recently, it has gained great attention in biomedical sciences. Based on the molecular imaging, molecular profiles and/or cell behavior altered prior to these alteration can be visualized anatomically. Thereby, this had predisposed molecular imaging to early detection of disease with high sensitivity and specificity.^[1,3]

Nanoparticles have gained great attention for applications in medicine, including cancer treatment and molecular imaging.^[4,5] There are different methods of molecular imaging, including magnetic resonance imaging (MRI),^[6] computed tomography (CT),^[7] ultrasound, and nuclear

medicine modalities.^[8-10] The important point is that each modality has limitations and advantages over each other, that is, none of them is able to completely give anatomical and pathological data independently.^[11] Therefore, it can be useful to combine two or more imaging modalities.

Among them, MRI is a noninvasive and nonionizing type of radiation with high spatial resolution and superior soft-tissue contrast.^[12,13] Meanwhile, CT imaging has a high resolution, three-dimensional tomography technique, so that its density resolution is better than alternative imaging modalities.^[14] Therefore, combining CT and MRI can help to accurate disease diagnosis.

Nanoparticles have gained great attention for applications in molecular imaging. In recent decades, superparamagnetic

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Mohammad Keshtkar¹,
Daryoush Shahbazi-Gahrouei²,
Alireza Mahmoudabadi¹

¹Department of Medical Physics and Radiology, Gonabad University of Medical Sciences, Gonabad, ²Department of Medical Physics, Isfahan University of Medical Sciences, Isfahan, Iran

Address for correspondence:
Dr. Mohammad Keshtkar,
Department of Medical Physics and Radiology,
Gonabad University of Medical Sciences, Gonabad, Iran.
E-mail: keshtkar.dmohammad@yahoo.com

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iron oxide nanoparticles have been typically utilized as T₂-weighted MRI contrast agent due to their capability to dephase transverse magnetization and thereby reducing T₂ relaxation times of water protons.^[15] Nanoparticles with high X-ray absorption can be utilized as a contrast agent for CT imaging. For example, gold nanoparticles due to their higher X-ray absorption coefficient and good biocompatibility have been widely applied for CT imaging.^[16] Therefore, combination of Fe₃O₄ and Au nanoparticles have been applied for MRI/CT dual-modality imaging.^[17]

There are some studies about synthesizing and application of different nanocomposites with different construction method in dual-modal imaging, such as polyethylenimine (PEI)-entrapped gold nanoparticles chelated with gadolinium (Gd) ions,^[18] TbF₃,^[19] GdF₃,^[20] strawberry-like Fe₃O₄-Au nanoparticles,^[21] gadolinium-loaded dendrimer-entrapped gold nanoparticles,^[22] PEI-Au-Gd nanoparticles,^[23] Fe₂O₃@Au nanoparticles,^[24] FePt,^[25] and Fe₃O₄/Au.^[26]

It has been proved that Fe₃O₄ nanoparticles can improve image contrast and sensitivity in MRI.^[26] Furthermore, gold improves CT image contrast in nanoparticle format.^[22] Yu *et al.*^[27] and Hu *et al.*^[11] developed dumbbell-like Au-Fe₃O₄ nanocomposites in heating condition of approximately 300°C and Fe₃O₄/Au nanocomposites at 80°C, respectively. He *et al.* developed Fe₂O₃@Au nanoparticles by oxidizing Fe₃O₄ seed using HNO₃ solution with stirring at 100°C.^[24] Cai *et al.* reported developing Fe₃O₄/Au nanocomposites in relatively high cost, complicated and time-consuming manner with layer by layer self-assembly technique and iterative Au salt reduction process.^[26] In the synthesizing process, it is desirable to develop nanocomposites in relatively easy manner, using low-cost materials and in-room temperature; therefore, the main goal of this manuscript is to evaluate new developed nanocomposite structure for magnetic resonance (MR)/CT imaging.

Materials and Methods

Materials

Ferrous chloride tetrahydrate (FeCl₂·4H₂O, >99%), ferric chloride hexahydrate (FeCl₃·6H₂O) (>99%), hydrochloric acid (HCl, 37%), Si(OC₂H₅)₄ (tetraethyl orthosilicate [TEOS]), ammonia aqueous (25 wt%), and ethanol and toluene (>99%) were purchased from Merck. Sodium borohydride (NaBH₄), 3-aminopropyltriethoxysilane (APTES), and chloroauric acid (HAuCl₄) were obtained from Sigma (St. Louis, MO, USA). Deionized water was used throughout the experiment.

Synthesis of Fe₃O₄@Au nanoparticles

Chemical co-precipitation of Fe²⁺ and Fe³⁺ salts by the addition of a base in aqueous media was used for Fe₃O₄ synthesis.^[28] First, FeCl₃·6H₂O, and FeCl₂·4H₂O with

certain molar ratio were dissolved in 40 ml HCl (0.4 M). Then, a 400 mL NH₃ solution was quickly added under vigorous stirring. After precipitation, Fe₃O₄ nanoparticles magnetically collected, repeatedly washed, and re-dispersed in 150 mL of deionized water.

Fe₃O₄ nanoparticles were coated with the SiO₂ by modifying Stober method.^[29] Briefly, 30 mL suspension of Fe₃O₄ nanoparticles, 15 mL ammonia, 30 mL water, and 1.9 mL TEOS were added into 300 mL 2-propanol under vigorous stirring for 18 h. Finally, Fe₃O₄@SiO₂ was collected by magnet separation and washed several times.

In the next step, the previous product was amino-functionalization by APTES. Briefly, 6 gr Fe₃O₄@SiO₂ and 16 mL APTES were added into 60 mL anhydrous toluene with stirring overnight while the solution was deoxygenized by the argon. The Fe₃O₄@SiO₂/NH₂ washed with acetone and finally dried under vacuum.

In the final stage, 30 mg Fe₃O₄@SiO₂/NH₂ was dispersed in 45 mL water at pH = 4 under ultrasonication for 60 min. Subsequently, 15 mL HAuCl₄ aqueous solution (1.71 mM) was added to the MNPs. At the end, 15 mL NaBH₄ solution (0.1 M) was added to the mixture. Finally, the product was collected and washed using phosphate-buffered saline (PBS) (pH = 7.4).

Characterization

Transmission electron microscopy images were recorded on a Philips-EM208S at an accelerating voltage of 100 kV. Fourier transform infrared (FTIR) spectroscopy study was carried out by JASCO, FT/IR-6300 (Japan), to determine the chemical functional groups in the nanoparticles at various steps of synthesis. The magnetic properties of nanoparticles were evaluated by the Alternating Gradient Field Magnetometry at room temperature up to 9000 Oe. X-ray diffraction (XRD) study was through a Bruker X-ray diffractometer. The iron and gold concentrations of the stock solution were measured using an atomic absorption spectrophotometer (Shimadzu, AA-680).

Cell culture

The human hepatocellular carcinoma (HepG2) cells were grown in RPMI-1640 medium supplemented with 10% fetal bovine serum, penicillin (100 U/mL), and streptomycin (100 µg/mL). The cells were maintained at 37°C in a humidified incubator with 5% CO₂.^[30]

Cytotoxicity assay

In vitro cytotoxicity of the Fe₃O₄@Au nanoparticles was evaluated by 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay on HepG2 cells. Briefly, HepG2 cells were grown into 96-well cell plates at a density of 5,000 cells per well and incubated overnight in a humidified incubator with a CO₂ concentration of 5%. Then, the cells

were incubated with various concentrations of Fe₃O₄@Au nanoparticles ranging from 10 to 80 µg/mL for 24 h. A culture medium without any nanoparticles prepared as the control wells.

Thereafter, MTT (20 µL in PBS, 5 mg/mL) was added to each well and further incubated for 4 h. Then, the culture medium was carefully removed, and 200 µL DMSO was added to each well to dissolve insoluble Formazan crystals. The absorbance at 570 nm in each well was recorded using an ELISA reader (Synergy H1, Bio Tek). Mean and standard deviation of three wells were expressed for each sample. The percentage of cell viability was measured by dividing viable cells in each well by control wells.

T₂ relaxivity measurement and X-ray attenuation property

The efficiency of an MRI contrast agent is determined by its ability to manipulate relaxation rates.^[31] Due to MRI contrast agents change the T₁ and T₂ relaxation times of the abundant water, the relaxivity of a contrast agent is presented by its change in the water proton relaxation times.^[32] In the case of T₂-weighted contrast agent, transverse relaxation rates are altered in the existence of a contrast agent by

$$R_2 = 1/T_2 + r_2 C. [32]$$

Where C is its concentration, and r₂ is transverse relaxivity. Moreover, in the absence of contrast agents, C = 0, R₂ is unperturbed water transverse relaxation rate.

For determining relaxivity, solutions of Fe₃O₄@Au nanoparticles were prepared in water at different iron concentrations. MRI was performed at room temperature using a 1.5 T scanner (Symphony, Siemens). T₂ relaxivity was determined using multiple spin-echo sequences utilizing 16 echo-times ranging from 22 to 352 ms and a repetition time of 3000 ms.^[33] The other parameters were set as follow: slice thickness = 3 mm, number of excitation = 1, acquisition matrix = 128 × 128. T₂ relaxivity (r₂ in mM⁻¹s⁻¹) was calculated from the slopes of 1/T₂ against Fe concentration.

Evaluating the ability of the Fe₃O₄@Au nanoparticles in attenuating X-ray was performed using a clinical 64 slice scanner (SOMATOM sensation, Siemens) with 80 kV, 300 mAs, and a slice thickness of 0.6 mm. The samples prepared in water with various Au concentrations were put into 2 mL Eppendorf tubes. The assessment of nanoparticles in X-ray attenuation was performed by importing the CT images in ImageJ program and then drawing a region of interest in CT image for each sample.

Results

Nanoparticles characterization

The size of the Fe₃O₄@Au nanoparticles was revealed by TEM imaging. The obtained Fe₃O₄@Au nanoparticles had a spherical morphology with a size of <75 nm [Figure 1].

The structure of the synthesized Fe₃O₄@Au nanoparticles was characterized by XRD [Figure 2]. Theta is the angle between incident and reflected ray. The diffraction peaks observed at 30.3° (220), 35.6° (311), 43.2° (400), 53.4° (422), 57.2° (511), 62.7° (440) could be assigned to Fe₃O₄. Furthermore, the peaks observed at 38.2°, 44.4°, 64.5°, and 77.4° could be related to the reflections of the (111), (200), (220), and (311) crystalline planes of cubic Au, respectively.

The chemical composition of the obtained nanoparticles was analyzed by FTIR spectrum as shown in Figure 3. The absorption peaks at 3417 are associated with the N–H stretching vibration. Furthermore, the presence of the propyl groups in APTES molecules is revealed by C–H stretching vibrations that emerged at 2930 and 2862 cm⁻¹. It should be noted that there is no absorption band for the Au in the infrared region.

Figure 4 shows the magnetic hysteresis loop of Fe₃O₄@Au nanoparticles at room temperature. In fact, it represents the plot of the magnetization “M” versus the applied field “H” (between –9 and +9 kOe) of the synthesized Fe₃O₄@Au nanoparticles. The saturation magnetization (M_s) was around 33 emu/g.

Cytotoxic assay

The biocompatibility of the nanoparticles was assessed on HepG2 cells by MTT assay. Figure 5 showed the result of MTT assay at different concentrations. The results are presented in terms of percentage cell viability. Materials with cell viability above 80% can be accepted as biocompatible material.^[34] Findings showed that for HepG2 cells, at concentration of 80 µg/mL Fe₃O₄@Au nanoparticles have cytotoxicity effects. Indeed, at concentrations from 10 up to 60 µg/mL cell toxicity is low or moderate.

T₂ relaxivity measurement and X-ray attenuation property

The role of Fe₃O₄ nanoparticles as an MRI contrast agent is to decrease transverse relaxation time. T₂-weighted MR

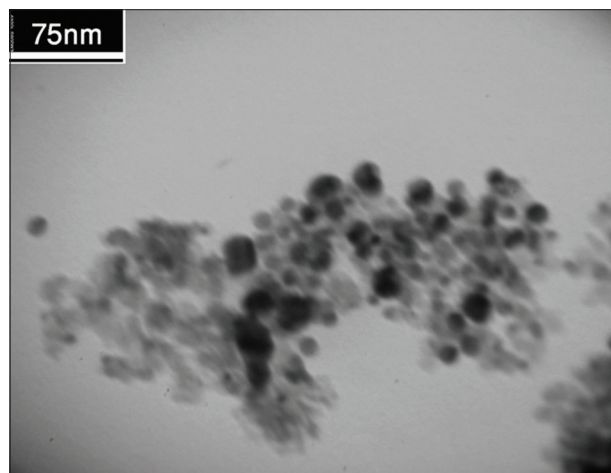


Figure 1: Transmission electron microscopy image of prepared Fe₃O₄@Au nanoparticles

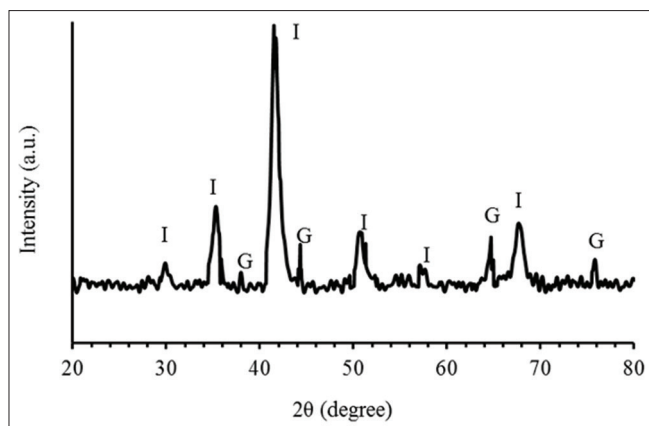


Figure 2: X-ray diffraction pattern of the formed Fe₃O₄@Au nanoparticles ("I" and "G" for Fe₃O₄ and Au nanoparticles, respectively)

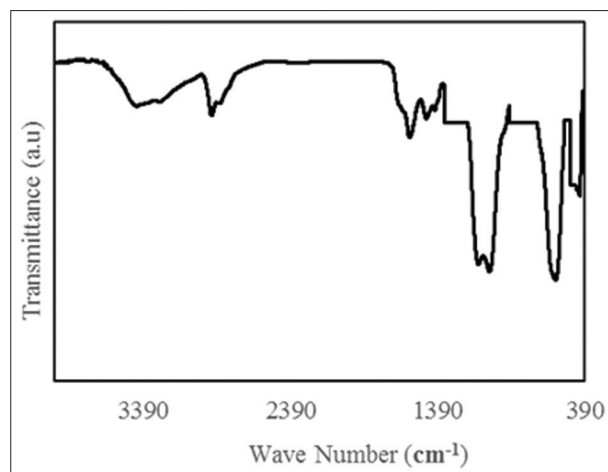


Figure 3: Fourier transform infrared spectrum of Fe₃O₄@Au nanoparticles

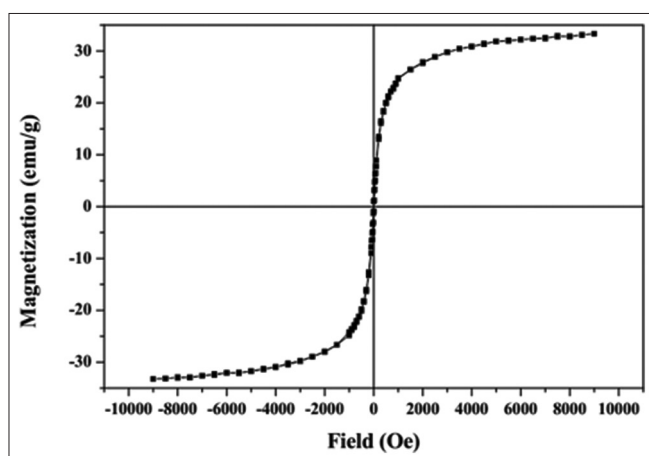


Figure 4: Magnetization curve of Fe₃O₄@Au nanoparticles

relaxometry of the Fe₃O₄@Au nanoparticles was performed to confirm their application as a contrast agent for MRI [Figure 6a]. Transverse relaxivity (r_2) was measured by plotting $1/T_2$ against the iron concentration in the solution [Figure 6b]. The measured relaxivity, which is the slope of the plot was $135.26 \text{ mM}^{-1}\text{s}^{-1}$.

X-ray attenuation property of Fe₃O₄@Au nanoparticles was investigated to confirm their use as CT contrast agent. Figure 7 shows the CT image of nanoparticles with different Au concentrations (a) and the plot of the Hounsfield units against Au concentration (b). As the Au concentration increases, the CT image intensity increases.

Discussion

Figure 1 shows the TEM image of Fe₃O₄@Au nanoparticles, and it reveals that the size of nanoparticles is $<70 \text{ nm}$. TEM image also shows some aggregated particles, which could be attributed to the TEM sample preparation process.

XRD analysis represents that the pattern of Fe₃O₄@Au nanoparticles coincides with reference pattern for magnetite and gold. The XRD results suggest the presence of both Fe₃O₄ and Au crystals in the prepared sample. XRD

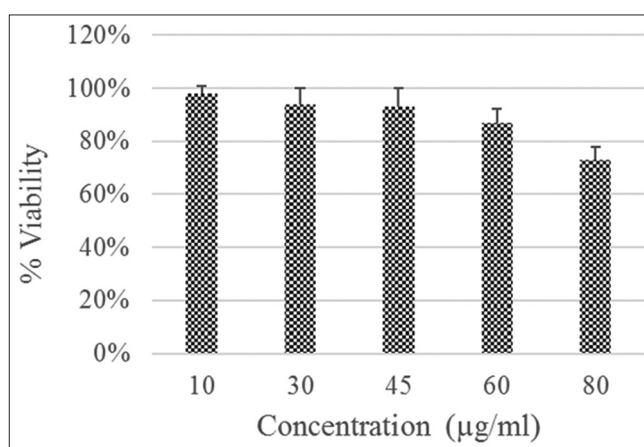


Figure 5: 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide assay of human hepatocellular carcinoma cells after incubation with Fe₃O₄@Au nanoparticles at different iron concentrations (10–80 µg/mL) after 24 h

provides information about structure and morphology of samples.

According to the FTIR spectrum, the presence of Fe₃O₄ nanoparticles can be seen by the absorption peak at 592 cm^{-1} , which is related to the Fe–O of magnetite phase. The adsorption of SiO₂ layer onto the surface of Fe₃O₄ was confirmed by bands at $1094, 957, 804,$ and 471 cm^{-1} which were ascribed to the formation of silica shells on the surface of Fe₃O₄. Meanwhile, there is no absorption band for the Au in the infrared region.

According to the magnetometry result [Figure 4], it was revealed that the nanoparticles had strong magnetic responses to a changing magnetic field. Little hysteresis of nanoparticles implies that they were possess superparamagnetic properties at room temperature.^[35]

MTT assay results confirmed that prepared Fe₃O₄@Au nanoparticles had little cytotoxicity up to 60 µg/mL and can be used safely for biomedical applications. Montazerabadi *et al.* synthesized Fe₃O₄@Au nanoparticles, and MTT assay showed that their nanoparticles did not have any

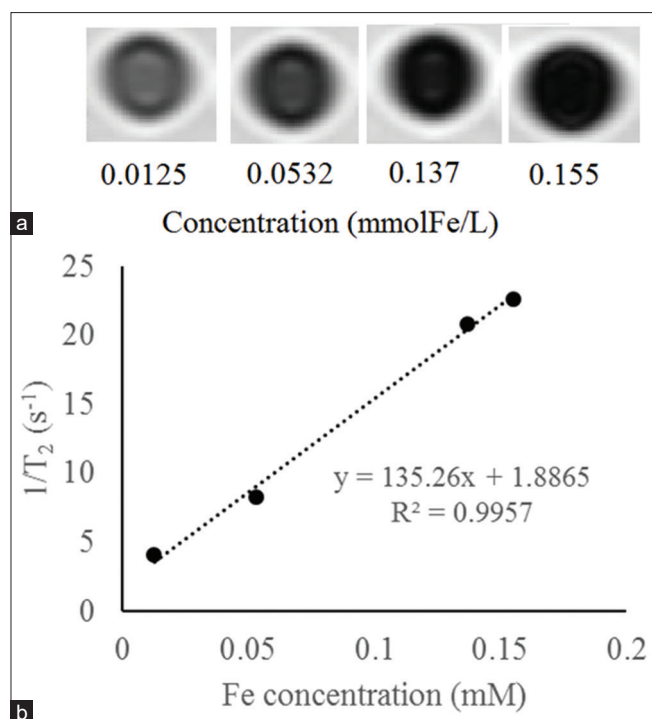


Figure 6: Magnetic resonance image (a) and linear fitting of transverse relaxivity rate (b) of the Fe₃O₄@Au nanoparticles with various Fe concentrations

cytotoxic effect on LNCaP cells.^[36] The highest used Fe concentration was 115 μg/mL. Their nanoparticles showed better cytocompatibility.

Figure 6a shows that increasing in Fe concentration, decreases the signal intensity. Moreover, the MR relaxometry revealed that transverse relaxivity was 135.26 mM⁻¹s⁻¹. These findings confirmed the capability of synthesized Fe₃O₄@Au nanoparticles as an MRI contrast agent.

Figure 7 shows that as the Au concentration increases, the signal intensity increases linearly. The results were demonstrated the potential of Fe₃O₄@Au nanoparticles for CT imaging.

There are several preparation methods for Fe₃O₄/Au nanoparticles. For example, by decomposing iron pentacarbonyl on the surfaces of Au nanoparticles can prepare dumbbell-like Fe₃O₄/Au nanoparticles.^[27] Furthermore, core/shell Fe₃O₄/Au nanoparticles can be prepared by synthesizing Au and thiol-functionalized nanoparticles separately and linking them by chemical bonds.^[37] Another way is assembling negatively charged Au nanoparticles onto the surface of the positively charged Fe₃O₄ nanoparticles.^[38] However, in this study, magnetic iron oxide nanoparticles were prepared by co-precipitation method and then were coated with silica according to Stober method. Consequently, silica-coated nanoparticles were amino-functionalized. Finally, gold nanoparticles assembled onto the surfaces of the previous product. In this work, the synthesizing process accomplished all at

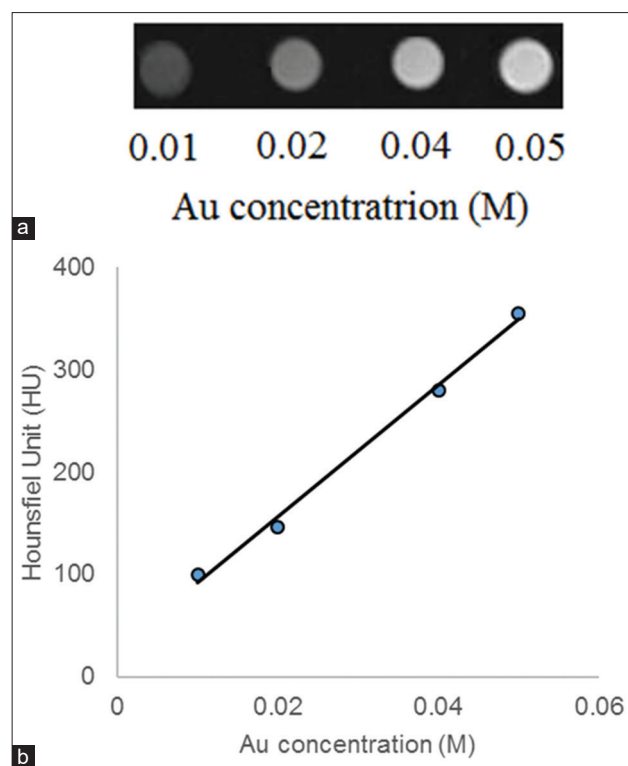


Figure 7: Computed tomography image of the Fe₃O₄@Au nanoparticles with different Au concentration (a) and X-ray attenuation of the Fe₃O₄@Au nanoparticles against Au concentration (b)

room temperature using convenient manner and relatively low-cost materials.

Because the main result of developing dual-modal contrast agent is to evaluate their application in imaging modalities (here, MRI and CT), their relaxivity and X-ray attenuation should be discussed. The results of the present work are in agreement with Hu *et al.*, where they synthesized hyaluronic acid-modified Fe₃O₄/Au nanocomposite as targeted contrast agent for CT and MRI. Their results showed that the formed nanoparticles represent a high r_2 relaxivity (264.16 mM⁻¹s⁻¹) and good X-ray attenuation property.

Li *et al.*^[39] reported synthesizing Fe₃O₄@Au-mPEG-PEI. NH₂ composite nanoparticles for dual-mode MR/CT imaging. Their results showed that nanoparticles had a relatively high r_2 relaxivity of 146.07 mM⁻¹s⁻¹ and good X-ray attenuation. The results of our study are in good agreement with Li *et al.* findings.

Cai *et al.*^[40] introduced Fe₃O₄/Au nanocomposites by multilayer of PGA/PLL/PGA. They results showed that uncoated Fe₃O₄, Fe₃O₄@G5, and Fe₃O₄@Au presented r_2 relaxivity of 277.81, 68.98, and 71.55 mM⁻¹s⁻¹, respectively. This decreasing in the value of relaxivity may be related to multilayer coating of Fe₃O₄ nanoparticles.

In another study, Cai *et al.*^[26] reported that synthesized Fe₃O₄/Au nanocomposites assisted by dendrimer and

using layers of poly glutamic acid and polylysine have r_2 relaxivity of 92.67 mM⁻¹s⁻¹ and good X-ray attenuation. Lower relaxivity compared to our study may be related to more layers on Fe₃O₄ nanoparticles.

TbF₃ nanoparticles developed by Zheng *et al.*^[19] showed a high transverse relaxivity of 395.77 mM⁻¹s⁻¹ under 7T magnetic field. This higher relaxivity may be attributed to high magnetic field.

By considering the use of 1.5 T MRI and multicoating of Fe₃O₄ nanoparticles in this study, our results are in good agreement with other similar researches. In addition, the findings of the present study suggested that developed Fe₃O₄@Au nanoparticles in easy and relatively low-cost manner can be utilized effectively in both CT and MRI applications.

Conclusion

In summary, Fe₃O₄@Au nanoparticles were developed as a dual-modality MR/CT imaging contrast media. Fe₃O₄ nanoparticles by shortening transverse relaxation time and Au nanoparticles by high X-ray attenuation property can be used for both MRI and CT, respectively. Fe₃O₄@Au nanoparticles showed good cytocompatibility up to 60 µg/mL, good r_2 relaxivity of 135.26 mM⁻¹s⁻¹ and a good X-ray attenuation property. It is concluded that Fe₃O₄@Au nanoparticles are able to be used as a contrast agent for dual-mode MR/CT imaging.

Conflicts of interest

There are no conflicts of interest.

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BIOGRAPHIES



Mohammad Keshtkar is an assistant professor of Medical Physics at the Department of Medical Physics and Radiology, Gonabad University of Medical Sciences, Gonabad, Iran. He received his B.Sc. in Radiology Technology from School of Paramedical Sciences, Shiraz University of Medical Sciences, Shiraz, Iran, M.Sc. degree

in Medical Physics from Tehran University of Medical Sciences, Tehran, Iran and Ph.D. in Medical Physics from Isfahan University of Medical Sciences, Isfahan, Iran.

Email: keshtkar.dmohammad@yahoo.com



Daryoush Shahbazi-Gahrouei is a Professor of Medical Physics at the Isfahan University of Medical Sciences, Isfahan, Iran. He received his B.Sc. in Physics from School of Science, Isfahan University, Iran, M.Sc. degree in Medical Physics from Tarbiat Modarres University, Tehran, Iran, and Ph.D. in Medical Physics from University of

Western Sydney and St. George Cancer Care Centre, Sydney, Australia.

Email: shahbazi@med.mui.ac.ir



Alireza Mahmoudabadi is an assistant professor of Radiology at the at the Department of Medical Physics and Radiology, Gonabad University of Medical Sciences, Gonabad, Iran. He received his MD degree from School of Medicine, Mashhad University of Medical Sciences, Mashhad and Radiology from School of

Medicine, Mashhad University of Medical Sciences, Mashhad, Iran.

Email: Mahmoudabadi_md@yahoo.com